nucleotide sequence and/or amino acid sequence disclosures pursuant to 37 CFR §1.821-1.825. The new sequences contained in the substitute Sequence Listing were present in the application as filed but were not assigned a sequence identification number. The foregoing amendments to the specification insert sequence identification numbers within the specification as required by 37 C.F.R. 1.821(d). The amendment adds no new matter because all sequences added to the sequence listing are present and find support in the application as filed.

I hereby state that the contents of the paper and computer readable copies of the substitute Sequence Listing, submitted herewith in accordance with 37 C.F.R. § 1.825(a) and (b) are the same.

This response is timely filed with a one month extension of time. Enclosed is a copy of the Notice together with our check in the amount of $240.00 ($130.00 missing parts surcharge and $110.00 extension of time fee). The Notice to File Missing Parts was mailed without a copy of the Raw Sequence Listing Error Report and therefore the Applicants were unable to submit this response within the required two month time period.

The Commissioner is hereby authorized to charge any deficiency in the amount enclosed or any additional fees which may be required under 37 CFR 1.16 or 1.17 to Deposit Account No. 13-2855. A copy of this request is enclosed.

Please refund any overpayment to Marshall, O'Toole, Gerstein, Murray & Borun at the address below.

Respectfully submitted,

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April 30, 2001

EXHIBIT A

MARKED UP VERSION OF AMENDMENTS TO SPECIFICATION

Page 16 line 21:

In a preferred embodiment, the di-lysine-modified APP of the invention is a human APP. For example, human APP isoforms such as APP695, APP751, and APP770, modified to include the two lysines, are contemplated. In a preferred embodiment, the APP isoform comprises at least one variation selected from the group consisting of a Swedish KM→NL mutation and a London V717→F mutation, or any other mutation that has been observed in a subpopulation that is particularly prone to development of Alzheimer's disease. These mutations are recognized as mutations that influence APP processing into Aβ. In a highly preferred embodiment, the APP protein or fragment thereof comprises the APP-Sw β-secretase peptide sequence NLDA **(SEQ ID NO: 76)**, which is associated with increased levels of Aβ processing and therefore is particularly useful in assays relating to Alzheimer's research. More particularly, the APP protein or fragment thereof preferably comprises the APP-Sw β-secretase peptide sequence SEVNLDAEFR

Page 18 line 1:

In another, related embodiment, the invention provides a polypeptide useful for assaying for modulators of β-secretase activity, said polypeptide comprising an amino acid sequence of the formula NH_2-X-Y-Z-KK-COOH; wherein X, Y, and Z each comprise an amino acid sequence of at least one amino acid; wherein-NH_2-X comprises an amino-terminal amino acid sequence having at least one amino acid residue; wherein Y comprises an amino acid sequence of a β-secretase recognition site of a mammalian amyloid protein precursor (APP); and wherein Z-KK-COOH comprises a carboxy-terminal amino acid sequence ending in two lysine (K) residues. In one preferred variation, the carboxyl-terminal amino acid sequence Z includes a hyrdrophobic domain that is a transmembrane domain in host cells that express the polypeptide. Host cells that express such a polypeptide are particularly useful in assays described herein for identifying modulators of APP processing. In another preferred variation, the amino-terminal amino acid sequence X includes an amino acid sequence of a

reporter or marker protein, as described above. In still another preferred variation, the β-secretase recognition site Y comprises the human APP-Sw β-secretase peptide sequence NLDA **(SEQ ID NO: 76)**. It will be apparent that these preferred variations are not mutually exclusive of each other -- they may be combined in a single polypeptide. The invention further provides a polynucleotide comprising a nucleotide sequence that encodes such polypeptides, vectors which comprise such polynucleotides, and host cells which comprises such vectors, polynucleotides, and/or polypeptides.

Page 20 line 12:

The β-secretase cleavage site in APP is known, and it will be appreciated that the assays of the invention can be performed with either intact APP or fragments or analogs of APP that retain the β-secretase recognition and cleavage site. Thus, in one variation, the substrate polypeptide of the second composition comprises the amino acid sequence SEVNLDAEFR **(SEQ ID NO: 63)**, which includes the β-secretase recognition site of human APP that contains the "Swiss" mutation. In another variation, the substrate polypeptide of the second composition comprises the amino acid sequence EVKMDAEF **(SEQ ID NO: 70)**. In another variation, the second composition comprises a polypeptide having an amino acid sequence of a human amyloid precursor protein (APP). For example, the human amyloid precursor protein is selected from the group consisting of: APP695, APP751, and APP770. Preferably, the human amyloid precursor protein (irrespective of isoform selected) includes at least on mutation selected from a KM→NL Swiss mutation and a V→F London mutation. As explained elsewhere, one preferred embodiment involves a variation wherein the polypeptide having an amino acid sequence of a human APP further comprises an amino acid sequence comprising a marker sequence attached amino-terminal to the amino acid sequence of the human amyloid precursor protein. Preferably, the polypeptide having an amino acid sequence of a human APP further comprises two lysine residues attached to the carboxyl terminus of the amino acid sequence of the human APP. The assays can be performed in a cell free setting, using cell-free enzyme and cell-free substrate, or can be performed in a cell-based assay wherein the second composition comprises a eukaryotic cell that expresses amyloid precursor protein (APP) or a fragment thereof containing a β-secretase cleavage site. Preferably, the APP expressed by the host cell is an APP variant that includes two carboxyl-

terminal lysine residues. It will also be appreciated that the β-secretase enzyme can be an enzyme that is expressed on the surface of the same cells.

Page 30 line 2:

Sequence ID No. 4: Human Asp-2(a), predicted amino acid sequence. The Asp2(a) amino acid sequence includes a putative signal peptide comprising residues 1 to 21; and a putative pre-propeptide after the signal peptide that extends through residue 45 (as assessed by processing observed of recombinant Asp2(a) in CHO cells), and a putative propeptide that may extend to at least about residue 57, based on the observation of an observed GRR↓GS **<u>(SEQ ID NO: 77)</u>** sequence which has characteristics of a protease recognition sequence. The Asp2(a) further includes a transmembrane domain comprising residues 455-477, a cytoplasmic domain comprising residues 478-501, and a putative alpha-helical spacer region, comprising residues 420-454, believed to be unnecessary for proteolytic activity, between the protease catalytic domain and the transmembrane domain.

Page 33 line 25:

Figure 5: Figure 5 shows the BestFit alignment of the predicted amino acid sequences of Hu-Asp2(a) **<u>(SEQ ID NO: 4)</u>** and murine Asp2(a) **<u>(SEQ ID NO: 8).</u>**

Page 37 line 1:

Alignment of the amino acid sequences of Hu-Asp2 with other known aspartyl proteases reveals a similar domain organization. All of the sequences contain a signal sequence followed by a pro-segment and the catalytic domain containing 2 copies of the aspartyl protease active site motif (DTG/DSG) separated by approximately 180 amino acid residues. Comparison of the processing site for proteolytic removal of the pro-segment in the mature forms of pepsin A, pepsin C, cathepsin D, cathepsin E and renin reveals that the mature forms of these enzymes contain between 31-35 amino acid residues upstream of the first DTG motif. Inspection of this region in the Hu-Asp-2 amino acid sequence indicates a preferred processing site within the sequence GRR↓GS **<u>(SEQ ID NO: 77)</u>** as proteolytic processing of pro-protein precursors commonly occurs at site following dibasic amino acid pairs (*eg.* RR). Also, processing at this site would yield a mature enzyme with 35 amino acid residues upstream of the first DTG, consistent with the processing sites for other aspartyl

proteases. In the absence of self-activation of Hu-Asp2 or a knowledge of the endogenous protease that processes Hu-Asp2 at this site, a recombinant form was engineered by introducing a recognition site for the PreSission protease (LEVLFQ↓GP**; SEQ ID NO: 62**) into the expression plasmids for bacterial, insect cell, and mammalian cell expression of pro-Hu-Asp2. In each case, the Gly residue in P1' position corresponds to the Gly residue 35 amino acids upstream of the first DTG motif in Hu-Asp2.

Page 40 line 1:

Also provided herein are purified Hu-Asp polypeptides, both recombinant and non-recombinant. Most importantly, methods to produce Hu-Asp2 polypeptides in active form are provided. These include production of Hu-Asp2 polypeptides and variants thereof in bacterial cells, insect cells, and mammalian cells, also in forms that allow secretion of the Hu-Asp2 polypeptide from bacterial, insect or mammalian cells into the culture medium, also methods to produce variants of Hu-Asp2 polypeptide incorporating amino acid tags that facilitate subsequent purification. In a preferred embodiment of the invention the Hu-Asp2 polypeptide is converted to a proteolytically active form either in transformed cells or after purification and cleavage by a second protease in a cell-free system, such active forms of the Hu-Asp2 polypeptide beginning with the N-terminal sequence TQHGIR **(SEQ ID NO: 78)** or ETDEEP **(SEQ ID NO: 79)**. The sequence TQHGIR **(SEQ ID NO: 78)** represents the amino-terminus of Asp2(a) or Asp2(b) beginning with residue 22 of SEQ ID NO: 4 or 6, after cleavage of a putative 21 residue signal peptide. Recombinant Asp2(a) expressed in and purified from insect cells was observed to have this amino terminus, presumably as a result of cleavage by a signal peptidase. The sequence ETDEEP **(SEQ ID NO: 79)** represents the amino-terminus of Asp2(a) or Asp2(b) beginning with residue 46 of SEQ ID NO: 4 or 6, as observed when Asp2(a) has been recombinantly produced in CHO cells (presumably after cleavage by both a rodent signal peptidase and another rodent peptidase that removes a propeptide sequence). The Asp2(a) produced in the CHO cells possesses β-secretase activity, as described in greater detail in Examples 11 and 12. Variants and derivatives, including fragments, of Hu-Asp proteins having the native amino acid sequences given in SEQ ID Nos: 2, 4, and 6 that retain any of the biological activities of Hu-Asp are also within the scope of the present invention. Of course, one of ordinary skill in the art will readily be able to determine whether a variant, derivative, or fragment of a Hu-Asp protein displays Hu-Asp

activity by subjecting the variant, derivative, or fragment to a standard aspartyl protease assay. Fragments of Hu-Asp within the scope of this invention include those that contain the active site domain containing the amino acid sequence DTG, fragments that contain the active site domain amino acid sequence DSG, fragments containing both the DTG and DSG active site sequences, fragments in which the spacing of the DTG and DSG active site sequences has been lengthened, fragments in which the spacing has been shortened. Also within the scope of the invention are fragments of Hu-Asp in which the transmembrane domain has been removed to allow production of Hu-Asp2 in a soluble form. In another embodiment of the invention, the two halves of Hu-Asp2, each containing a single active site DTG or DSG sequence can be produced independently as recombinant polypeptides, then combined in solution where they reconstitute an active protease.

Page 50 line 11:

In another embodiment, the invention relates to a method of assaying Hu-Asp function, specifically Hu-Asp2 function which involves incubating in solution the Hu-Asp polypeptide with a suitable substrate including but not limited to a synthetic peptide containing the β-secretase cleavage site of APP, preferably one containing the mutation found in a Swedish kindred with inherited AD in which KM is changed to NL, such peptide comprising the sequence SEVNLDAEFR **(SEQ ID NO: 63)** in an acidic buffering solution, preferably an acidic buffering solution of pH5.5 (see Example 12) using cleavage of the peptide monitored by high performance liquid chromatography as a measure of Hu-Asp proteolytic activity. Preferred assays for proteolytic activity utilize internally quenched peptide assay substrates. Such suitable substrates include peptides which have attached a paired flurophore and quencher including but not limited to 7-amino-4-methyl coumarin and dinitrophenol, respectively, such that cleavage of the peptide by the Hu-Asp results in increased fluorescence due to physical separation of the flurophore and quencher. Other paired flurophores and quenchers include bodipy-tetramethylrhodamine and QSY-5 (Molecular Probes, Inc.). In a variant of this assay, biotin or another suitable tag may be placed on one end of the peptide to anchor the peptide to a substrate assay plate and a flurophore may be placed at the other end of the peptide. Useful flurophores include those listed above as well as Europium labels such as W8044 (EG&g Wallac, Inc.). Cleavage of the peptide by Asp2 will release the flurophore or other tag from the plate, allowing

compounds to be assayed for inhibition of Asp2 proteolytic cleavage as shown by an increase in retained fluorescence. Preferred colorimetric assays of Hu-Asp proteolytic activity utilize other suitable substrates that include the P2 and P1 amino acids comprising the recognition site for cleavage linked to o-nitrophenol through an amide linkage, such that cleavage by the Hu-Asp results in an increase in optical density after altering the assay buffer to alkaline pH.

Page 89 line 27:

Recombinant human Asp2(a) prepared in CHO cells and purified as described in Example 11 was used to assay Asp2(a) proteolytic activity directly. Activity assays for Asp2(a) were performed using synthetic peptide substrates containing either the wild-type APP β-secretase site (SEVKM↓DAEFR; **SEQ ID NO: 64**), the Swedish KM→NL mutation (SEVNL↓DAEFR; **SEQ ID NO: 63**), or the Aβ40 and 42 γ-secretase sites (RRGGVV↓IA↓TVIVGER; **SEQ ID NO: 65**). Reactions were performed in 50 mM 2-[N-morpholino]ethane-sulfonate ("Na-MES," pH 5.5) containing 1% β-octylglucoside, 70 mM peptide substrate, and recombinant Asp2(a) (1-5 μg protein) for various times at 37°C. The reaction products were quantified by RP-HPLC using a linear gradient from 0-70 B over 30 minutes (A=0.1% TFA in water, B=0.1%TFA/10%water/90%AcCN). The elution profile was monitored by absorbance at 214 nm. In preliminary experiments, the two product peaks which eluted before the intact peptide substrate, were confirmed to have the sequence DAEFR **(SEQ ID NO: 81)** and SEVNL **(SEQ ID NO: 82)** using both Edman sequencing and MADLI-TOF mass spectrometry. Percent hydrolysis of the peptide substrate was calculated by comparing the integrated peak areas for the two product peptides and the starting material derived from the absorbance at 214 nm. The sequence of cleavage/hydrolysis products was confirmed using Edman sequencing and MADLI-TOF mass spectrometry.

Page 91 line 3:

An alternative β-secretase assay utilizes internally quenched fluorescent substrates to monitor enzyme activity using fluorescence spectroscopy in a single sample or multiwell format. Each reaction contained 50 mM Na-MES (pH 5.5), peptide substrate MCA-EVKMDAEF[K-DNP] **(SEQ ID NO: 80)** (BioSource International) (50 μM) and purified Hu-Asp-2 enzyme. These components were equilibrated to 37 °C for various times

and the reaction initiated by addition of substrate. Excitation was performed at 330 nm and the reaction kinetics were monitored by measuring the fluorescence emission at 390 nm. To detect compounds that modulate Hu-Asp-2 activity, the test compounds were added during the preincubation phase of the reaction and the kinetics of the reaction monitored as described above. Activators are scored as compounds that increase the rate of appearance of fluorescence while inhibitors decrease the rate of appearance of fluorescence.

Page 98 line 6:

The relationship between Asp1 and APP processing was explored by determining if APP α-secretase, APP β-secretase, or APP γ-secretase peptide substrates were cleaved by recombinant Asp-1ΔTM(His)$_6$. These peptide substrates included the α-secretase specific substrates A$\beta_{10\text{-}20}$ and A$\beta_{12\text{-}28}$, the β-secretase specific substrates PHA-95812E (SEVKMDAEFR; SEQ ID NO: 64) and PHA-247574E (SEVNLDAEFR; SEQ ID NO: 63), and γ-secretase specific substrate PHA-179111E (RRGGVVIATVIVGER; SEQ ID NO: ~~76~~ 65). Each reaction consisted of incubating a peptide substrate (100 nM) with recombinant Asp-1ΔTM(His)$_6$ for 15 hours at pH 4.5 at 37°C. Reaction products were quantified by RP-HPLC at $A^{214\ nm}$. The elution profiles for Asp-1ΔTM(His)$_6$ were compared to those obtained from parallel Asp1 experiments. The identity of the cleavage products was determined by MADLI-TOF mass spectrometry. Table 6 summarizes the Asp1 substrates and indicates the cleavage site.